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Global patterns of functional trait variation along aridity gradients in bats

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Abstract

Aim: Our understanding of the biological strategies employed by species to cope with challenges posed by aridity is still limited. Despite being sensitive to water loss, bats successfully inhabit a wide range of arid lands. We here investigated how functional traits of bat assemblages vary along the global aridity gradient to identify traits that favour their persistence in arid environments.

Location: Global.

Time period: Contemporary.

Major taxa studied: Bats.

Methods: We mapped the assemblage-level averages of four key bat traits describing wing morphology, echolocation and body size, based on a grid of 100-km resolution and a pool of 915 bat species, and modelled them against aridity values. To support our results, we conducted analyses also at the species level to control for phylogenetic autocorrelation.

Results: At the assemblage level, we detected a rise in values of aspect ratio, wing loading and forearm length, and a decrease in echolocation frequency with increasing aridity. These patterns were consistent with trends detected at the species level for all traits.

Main conclusions: Our findings show that trait variation in bats is associated with the aridity gradient and suggest that greater mobility and larger body size are advantageous features in arid environments. Greater mobility favours bats' ability to track patchy and temporary resources, while the reduced surface-to-volume ratio associated with a larger body size is likely to reduce water stress by limiting cutaneous

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evaporation. These findings highlight the importance of extending attention from species-specific adaptations to broad scale and multispecies variation in traits when investigating the ability of species to withstand arid conditions.

KEYWORDS

aridity gradient, assemblage level, bats, body size, echolocation, functional trait variation, global patterns, wing morphology

1 | INTRODUCTION

Arid and semi-arid regions cover c. one third of the planet's land surface, encompassing key habitats for a wide array of species (Safriel et al., 2005). Additionally, the extent of such regions is predicted to increase by 11–23% by the end of the century, thus affecting a greater number of biological communities (Huang et al., 2016). Arid environments are characterized by water limitations and unpredictability in precipitation, which result in low primary productivity and large spatio-temporal variation in food resources (Maestre et al., 2016). These features, often in combination with high temperatures and other abiotic attributes associated with aridity, represent major stressors for local species, shaping assemblage composition (O'Connell & Hallett, 2019; Rymer et al., 2016; Schwimmer & Haim, 2009). A range of behavioural, morphological and physiological strategies and adaptations are employed by species to cope with these stressors. For example, vertebrates may reduce water stress in several manners, e.g. via increased skin resistance to cutaneous evaporative water loss, burrowing habits and high mobility to access scattered resources (Fuller et al., 2014; Oliveira & Scheffers, 2019; Rymer et al., 2016; Williams & Tieleman, 2005). These adaptations have been mostly studied in species inhabiting deserts, and we still have a poor understanding of which traits favour the presence of species with increasing aridity across broad geographical extents. Over large geographical scales, average functional traits of species assemblages are associated with environmental conditions along gradients as a result of the combination of environmental filtering and ecological adaptation (Barnagaud et al., 2017; Holt et al., 2018; Lawing et al., 2017; Violle et al., 2014). Describing variation of species traits along aridity gradients can, therefore, provide insights on which features contribute to the ability of species to withstand arid conditions (Rymer et al., 2016).

Bats are a diverse and ubiquitous order of mammals, being found across all continents, except the polar regions. Their widespread distribution is accompanied by a great morphological and ecological diversification, with the ability to occupy a wide range of environmental conditions, from temperate and tropical rain forests to hyper-arid deserts (Simmons, 2003). Bats are considered particularly sensitive to water loss due to the presence of naked wing membranes (Minnaar et al., 2014) and to the exposure to air convection during flight (Carpenter, 1986). Nonetheless, they are a successful group in arid environments (Carpenter, 1969). The nocturnal habits of bats represent an advantage to face aridity constraints, and species-specific adaptations to extreme aridity, such as reduced

active thermoregulation and modified kidney structure, further contribute to their high diversity in many arid ecosystems (Bondarenko et al., 2014; Carpenter, 1969; Munoz-Garcia et al., 2016). Thanks to these features, bats constitute a good candidate group for investigating functional traits' responses to increasing aridity.

Along the global spectrum of moist to arid environmental conditions inhabited by bats (hereafter 'aridity gradient'), a progressive reduction in primary productivity and vegetation cover occurs, alongside the increased pressure of the stressors mentioned above (Safriel et al., 2005). In this context, bats and other wildlife are gradually exposed to structurally more open environments. In echolocating bats, the primary functional traits that mediate their interaction with the structure of the environment relate to features of wing morphology and echolocation calls (Aldridge & Rautenbach, 1987). These features are associated, respectively, with flight abilities and sensory perception in relation to background objects. A high wing loading (high weight to wing surface ratio) confers a fast but poorly manoeuvrable flight, while high values of aspect ratio (narrow wings) are associated with energy-efficient flight (Norberg & Rayner, 1987). When it comes to echolocation, low frequency calls are less subject to atmospheric attenuation and can travel farther, being thus well suited to open spaces, whereas high frequency calls provide more detailed information about nearby objects and are associated with more cluttered environments (Denzinger & Schnitzler, 2013). Species foraging in open spaces (open space foragers) are typically adapted to fast flight, with high values of wing loading and aspect ratio, and tend to exhibit low echolocation frequencies (Norberg & Rayner, 1987). In contrast, bats flying within the vegetation (narrow space foragers) are characterized by manoeuvrable flight with low values of wing loading and aspect ratio and high echolocation frequencies (Denzinger & Schnitzler, 2013). With trait values covering a continuum between these two extremes, morphological and echolocation trait variation has been applied to identify the effects of an increase in open space following forest fragmentation, reporting the partial loss of narrow space foragers from deforested areas (Estrada-Villegas et al., 2010; Farneda et al., 2015; Hayes et al., 2007). Similarly, the progressive decrease in the structural complexity of vegetation with aridity might drive a shift in assemblage traits towards values associated with greater adaptation to open spaces.

The increase in aridity itself also exerts a greater water stress on species. Several adaptations to preserve water have been identified in bats, such as modified kidneys to concentrate urine, or torpor, which contributes to reducing respiratory evaporative water losses (Carpenter, 1969; Munoz-Garcia et al., 2016). Cutaneous evaporative

water loss, one of the significant routes of water loss in bats, is dependent on body surface area (Minnaar et al., 2014). It is possible that larger body sizes, characterized by a lower surface-to-volume ratio, are comparatively less exposed to water loss and thus favoured as aridity increases (Munoz-Garcia et al., 2016). This could be particularly true for bats, since larger bodied bats have relatively smaller wings when compared to smaller species (Findley et al., 1972), thus reducing exposure via the naked membranes of the wings. Alternatively, since aridity is often associated with warm temperatures, smaller sizes could be associated with high aridity as a feature favouring heat exchange, as stated by Bergmann's rule (Bergmann, 1847; Blackburn & Gaston, 1996; *sensu* Meiri, 2011; Torres-Romero et al., 2016). However, several studies suggested that this might not be generalizable across all taxonomic groups and geographical scales (Medina et al., 2007; Rodríguez et al., 2008). In echolocating bats (hereafter only 'bats'), the overall small body size across species might allow for sufficient heat dissipation in hot environments, while water retention might be a more critical constraint than temperature in highly arid regions. However, trends in traits identified along aridity and temperature gradients are likely not entirely comparable as a number of arid areas are located in cold regions (e.g. cold deserts).

In this study, we aimed to investigate the set of functional traits that enable bats to cope with arid conditions. We examined how a bat assemblage's measures of wing morphology (aspect ratio and wing loading), echolocation frequency and body size vary along the aridity gradient. Due to the decreasing vegetation structural complexity, we hypothesized that increasing aridity will be associated with bat assemblages with high wing loading and aspect ratio and low echolocation frequency. Due to the need to mitigate evaporative water loss, we also hypothesized that aridity will select for assemblages of bats with larger body sizes. To test our hypotheses, we conducted analyses at the assemblage level (with an assemblage defined as the set of echolocating bat species coexisting in the same grid cell). While a grid-based approach not accounting for phylogenetic relationships among species is appropriate to test ecological hypotheses (e.g. environmental filtering, de Bello et al. 2015), the inclusion of models incorporating phylogenetic signal can provide further information and disclose evolutionary patterns (de Bello et al. 2015). Therefore, we complemented our analysis by testing our hypotheses also at the species level via a phylogenetic comparative approach (Figure 1, see e.g. Olalla-Tárraga et al., 2010). In addition, this species-level

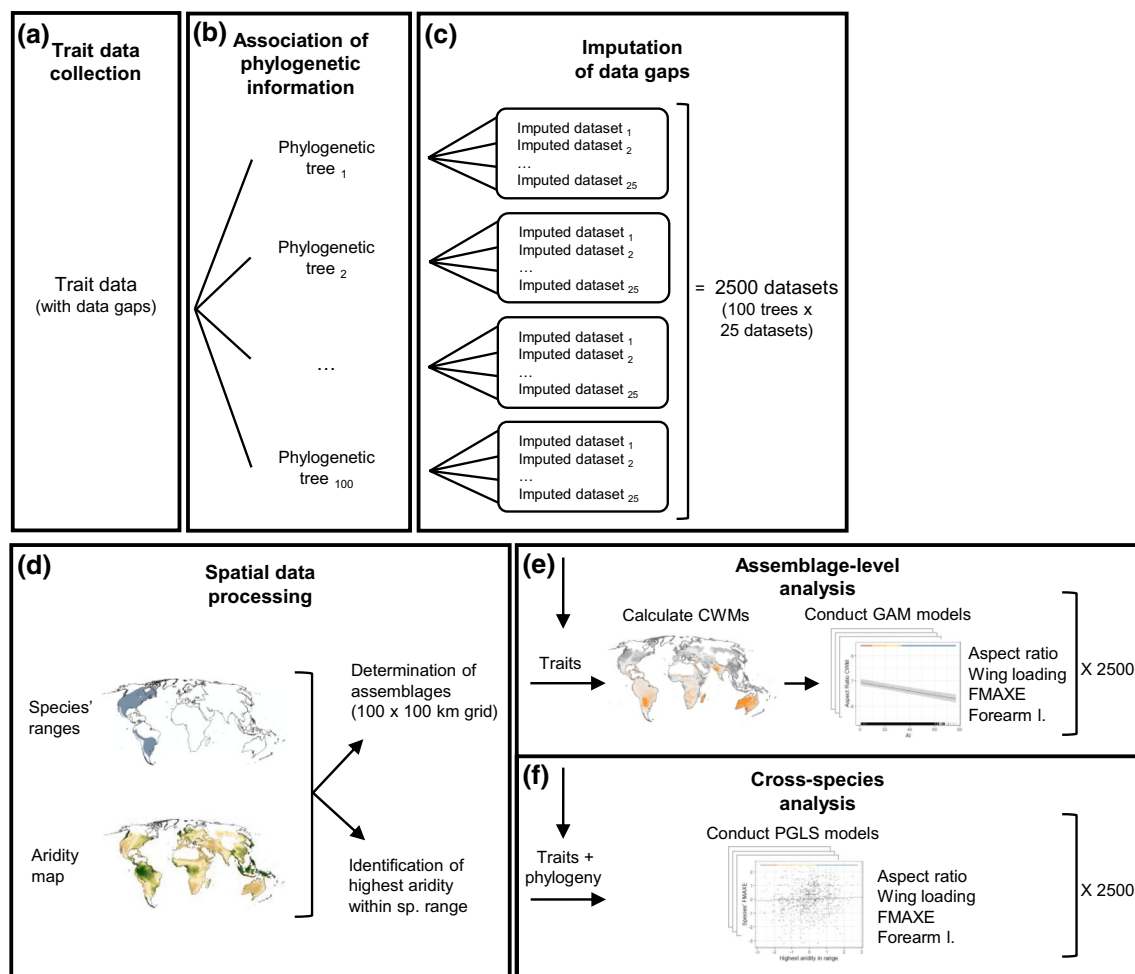


FIGURE 1 Diagram summarizing the main methodological steps. FMAXE = frequency of maximum energy; GAM = generalized additive model; CWM = community weighted mean; PGLS = phylogenetic generalized least squares [Colour figure can be viewed at wileyonlinelibrary.com]

analysis is not affected by spatial autocorrelation bias and, therefore, constitutes a suitable integration to our main analysis. The assemblage-level analysis required complete data, so we filled data gaps by means of multiple imputations. Phylogenetic information was employed both during data imputation, to improve imputation results, and when conducting the species-level analysis, to control for phylogenetic autocorrelation.

2 | MATERIALS AND METHODS

2.1 | Species data compilation and imputation

2.1.1 | Trait data

We obtained a list of bat species from the International Union for Conservation of Nature's (IUCN's) Red List of Threatened Species (IUCN, 2019), which was filtered to exclude bats of the family Pteropodidae, species for which distribution maps were not available ($n = 15$), and extinct species ($n = 1$). This led to a final sample of 1,058 species. We compiled species trait data using available mammal and bat trait databases (PanTHERIA, Jones et al., 2009; Phylacine, Faurby et al., 2018; EltonTraits, Wilman et al., 2014; EchoBank, Collen, 2012), as well as with targeted literature searches to reduce gaps in the dataset. A list of the data sources is found in the Appendix. We collected information on the main traits of interest, namely forearm length (as a proxy for body size), wing loading, aspect ratio and frequency of maximum energy (FMAXE) of the echolocation call (Figure 1a). Since it is recommended to impute data gaps using a larger set of correlated traits than those of interest to the study (Van Buuren, 2018), we also collated data on body mass and echolocation call duration. Wing loading and aspect ratio were defined following Norberg and Rayner (1987; see Supporting Information Appendix S1 for further details), while FMAXE was obtained for the harmonic with maximum energy (Monadjem et al., 2010). When collecting data from secondary sources (large databases) we discarded data obtained via extrapolation or other indirect methods.

2.1.2 | Imputations

We imputed data gaps using a multiple imputation technique (multiple imputation by chained equations, 'mice'; Van Buuren & Groothuis-Oudshoorn, 2011). A total of 915 bat species were included in the imputation model, after excluding species for which no trait data were available. Within this set of species, missing data represented 12% ($n = 110$) for forearm length, 60% ($n = 548$) for the wing loading, 59% ($n = 537$) for the aspect ratio and 44% ($n = 401$) for the FMAXE, while the auxiliary variables body mass and call duration contained, respectively, 13% ($n = 121$) and 44% ($n = 406$) missing data. The imputation model included all main traits plus body mass and call duration as auxiliary variables. Following Penone et al. (2014), we

also included phylogenetic relationships in the form of eigenvectors extracted from the phylogenetic distance matrix via a principal component analysis (Figure 1b). Phylogenetic information was missing for 1% ($n = 11$) of the species included in the imputation procedure. Phylogenetic relationships among species were based on the node-dated phylogeny of mammals developed by Upham et al. (2019). To account for phylogenetic uncertainty, we replicated imputations for 100 randomly selected trees. The first five eigenvectors represented on average c. 95% of the variance.

Data imputation can contribute to reducing sampling bias in available data (Nakagawa & Freckleton, 2008). However, imputed trait values may vary in their uncertainty depending on the proportion of missing data and on the correlation strength between the considered traits (Madley-Dowd et al., 2019). Multiple imputation generates multiple datasets and thus a distribution of possible trait values for each missing value, which reflects the uncertainty in the prediction. While it is common to only consider the mean or median of the distribution (e.g. Brum et al., 2017; Oliveira et al., 2016; Penone et al., 2014), this neglects the underlying uncertainty. Here, instead, we conducted the analyses on each of the imputed datasets, thus allowing the uncertainty of the imputation to propagate directly on the results (Cooke et al., 2019). As such, the uncertainty in the results reflects the uncertainty in the underlying data.

Imputation model parameters were set to produce 25 imputed datasets (optimized relative to the average percentage of missing data, 21.6%; Van Buuren, 2018), each generated via 50 iterations, and variable imputation order was set to maximize convergence (Van Buuren, 2018). The imputation procedure, therefore, produced 2,500 datasets, 25 for each of the 100 trees considered (Figure 1c). Imputation model convergence as well as quality of the results were assessed for 10 randomly selected trees of the ones in use via visual inspection of imputation outputs following Van Buuren (2018) (see Supporting Information Appendix S2 for further methodological details). We further checked imputation accuracy by introducing an additional artificial 5% of missing data to the original data and by verifying that imputed data approximates observed data. To compare observed and imputed data we employed the percent bias, which measures the average tendency of the imputed values to be larger or smaller than the observed ones (Zambrano-Bigiarini, 2017). For satisfactory performance, the upper accepted limit for the percent bias is 5% (Demirtas et al., 2008; Van Buuren, 2018). We conducted this for 10 randomly selected trees of the ones in use, and simulated data were compared to observed data after obtaining the median across the 25 imputed datasets. All 2,500 datasets were retained and used to assess the influence of imputation uncertainty on the results of both the assemblage- and species-level analyses.

2.2 | Spatial data processing

To estimate assemblage-level average traits, we generated a global grid of 100 km × 100 km resolution using the equal-area Mollweide projection, and for each cell determined species presence and cell

areas covered using IUCN range maps (Figure 1d). This resolution is often considered a good compromise to investigate broad scale geographical patterns while accounting for the intrinsic uncertainty in the range maps (Buckley & Jetz, 2008; Santini et al., 2017). For each imputed dataset, we then calculated the assemblage mean of each trait per cell, where each species value is weighted by the area of the cell that its range covers (often referred to as community weighted mean, hereafter only 'CWM'). CWMs were calculated using the function 'ddply' from the R package 'plyr' (Wickham, 2011). We excluded cells with fewer than five species ($n = 3,527$, 24.8% of the total of cells containing at least one bat) from the grid to avoid linking aridity levels with non-representative assemblage values and provide greater comparability across assemblages (see e.g. Rapacciuolo et al., 2017).

We obtained aridity values from the map of the Global Aridity Index at 30 arc-seconds (Trabucco & Zomer, 2019; Supporting Information Figure S3.9), which was rescaled at the 100 km \times 100 km resolution in order to assign a value of aridity to each assemblage. It is important to note that the Global Aridity Index (hereafter only 'AI') is expressed as the ratio between mean annual precipitation and mean annual evapotranspiration (Trabucco & Zomer, 2019). Therefore, the greater is the AI, the less arid is the environment. Since our focus is on the influence of aridity on traits, we will discuss trends referring to 'increase in aridity' and 'decrease in aridity', which correspond, respectively, to smaller and greater AI values.

Data for the species-level analysis were obtained by extracting the value of the 5th percentile of the AI (high aridity) within the species geographical ranges (Figure 1d). This allowed us to focus on the putative highest aridity level experienced by each species in its distribution.

2.3 | Assemblage-level analysis

To examine variation in community functional traits along the global aridity gradient while controlling for spatial autocorrelation using a trend surface analysis, we employed generalized additive models (GAMs, Wood, 2017). While aridity was included as a linear predictor, spatial patterns were modelled by including a two-dimensional smoother of the coordinates of the centroid of each cell (see e.g. Fletcher & Fortin, 2018; Safi et al., 2011). This also accounted for other unobserved variables potentially affecting trait variation spatially. We verified the efficacy of the GAMs in controlling the spatial autocorrelation of the data with the use of variograms (see Supporting Information Figure S4.10). Models were built independently for each trait of interest and they were run across all datasets produced via the imputations, for a total of 10,000 models, 2,500 for each trait (Figure 1e). To exclude potential outliers, each model was re-run after excluding the 0.1% most influential observations as identified by the function 'influence.gam' from the 'mgcv' R package (Wood, 2011). The models were specified as follows:

$$CWM \sim AI + s(X, Y)$$

where CWM represents the community weighted mean for a certain trait, AI is the AI value for the assemblage and $s(X, Y)$ specifies a two-dimensional spline smoother for the spatial position of the cells. Before fitting the models, the AI was raised to the power of 0.4 to improve normality. We chose a 0.4 power transformation as it proved to be the most effective in improving normality of the AI. GAMs were fit using the function 'gam' (Wood, 2011). We tested the significance of aridity for modelling each trait's CWM by comparing the full models as described above to null models retaining only the spatial component. If the difference in the Akaike information criterion (ΔAIC) between the null and full model was ≥ 2 , aridity was considered significant in explaining trait variation. We repeated all validation procedures for 25 randomly selected datasets out of 2,500. Results across models were summarized for each trait by providing medians and 95% confidence intervals of models' parameters.

To ensure that the trends observed were not an artefact of un-included factors, we calculated the Pearson's r for the correlations between the AI and other environmental variables that could potentially affect CWM, namely annual mean temperature, temperature seasonality and precipitation seasonality. We obtained these environmental layers from WorldClim version 2.1 (Fick & Hijmans, 2017) and processed them analogously to the aridity layer. In particular, we projected and rescaled them at the 100 km \times 100 km resolution, extracted values for the cells included in the analysis and then, calculated Pearson's r of the correlations with the AI.

2.4 | Species-level analysis

As an independent test of the robustness of our results, we also tested our hypotheses using species as the unit of the analysis and accounting for phylogenetic autocorrelation. We employed phylogenetic generalized least squares (PGLS) models to assess the variation of traits across species as a function of the aridity level tolerated by each species (Figure 1f). Using the 5th quantile instead of the mean or the median of the AI value allowed us to focus on the putative extreme aridity experienced by the species within their current distribution while excluding possible errors due to the coarseness of these maps. We also tested weather using the 2.5th quantile or the median would result in qualitatively different results (see Supporting Information Table S4.3). To facilitate model convergence, trait data were log-transformed and the AI was raised to the power of 0.4. Analogously to the assemblage-level analysis, we produced a total of 10,000 PGLS models, 2,500 for each trait, and each model was re-run after excluding model-specific outliers. Outliers were identified as observations with studentized residuals $> \pm 3$, following Jones and Purvis (1997). The models were fitted via the function 'glsl' from the R package 'nlme' (Pinheiro et al., 2019) by specifying a Pagel's λ correlation structure available from the package 'ape' (Pagel, 1999; Paradis & Schliep, 2019), and they included 904 species for which phylogenetic information was available (Figure 1f). Results across models were summarized for each trait by providing medians and 95% confidence intervals of models' parameters. We provide a

measure of the phylogenetic signal detected for the relationship of each trait with aridity via the Pagel's λ as estimated by the models.

All analyses were conducted in the R environment, version 3.6.1 (R Core Team, 2019).

3 | RESULTS

3.1 | Imputation quality assessment

An inspection of diagnostic graphs for imputation accuracy indicated that model convergence was reached after *c.* 20 iterations and illustrated close resemblance between distributions of observed and imputed data (see Supporting Information Figures S2.2–S2.7 for diagnostic graphs for a sample tree). The comparison between imputed and observed data for artificially added missing records always produced a percent bias below the threshold of 5% (Supporting Information Table S2.1), confirming the suitability of multiple imputation by chained equation to fill in data gaps in our dataset.

3.2 | Assemblage-level analysis

The assemblage-level analysis included 915 bat species distributed over a grid of 10,669 cells. Each cell contained between 5 and 129 species and covered an aridity gradient ranging from hyper-arid (min AI \sim 0) to humid conditions (max AI = 4.47), with 95% of the cells located between AI = 0.03 and AI = 2.20. Pearson's *r* indicated low levels of correlations between aridity and other environmental variables (see Supporting Information Figure S3.8).

From the geographical representation of the CWMs, we can identify major patterns in trait variation and regions retaining extreme values (Figure 2a–d). Globally, aspect ratio and wing loading peak within arid regions (Figure 2a,b, see also Supporting Information Figure S3.9 for representation of aridity classes). The highest FMAXE are observed in the humid Southeast Asia, with some high values also located in the arid Arabian Peninsula, while a low peak is observed both in the arid zone of Australia and South America and in the Nearctic and Palearctic zones (Figure 2c). Forearm length, instead, is greater in the semi-arid and arid regions of India and in the arid and hyper-arid zones surrounding the Sahara, with low values widespread in the Nearctic and Palearctic zones (Figure 2d).

Model selection indicated that the models including the AI (full models) performed substantially better than null models in explaining the trait data (Δ AIC always well above 2, see Supporting Information Table S4.2). As predicted, aspect ratio and wing loading (addressing mobility) showed a positive relationship with increasing aridity [across 2,500 models for each trait, aspect ratio: median β_{aridity} (95% Confidence Intervals (CIs)) = -0.0084 (-0.0104 – -0.0064), $R^2 = .62 \pm .02$; wing loading: -0.031 (-0.0371 – -0.0254), $R^2 = .62 \pm .02$], and FMAXE (echolocation call frequency of maximum energy) exhibited a decreasing pattern with aridity [0.1686 (0.1244 – 0.2094), $R^2 = .78 \pm .01$; Figure 3a and Supporting Information Figure S4.11]. Forearm length (a proxy for body size) increased with aridity following our predictions [-0.0633 (-0.067 – -0.0594), $R^2 = .60 \pm .01$; Figure 3a and Supporting Information Figure S4.11]. Distribution of coefficients across all models produced indicated aridity as important for all traits. The patterns presented are consistent across all imputed

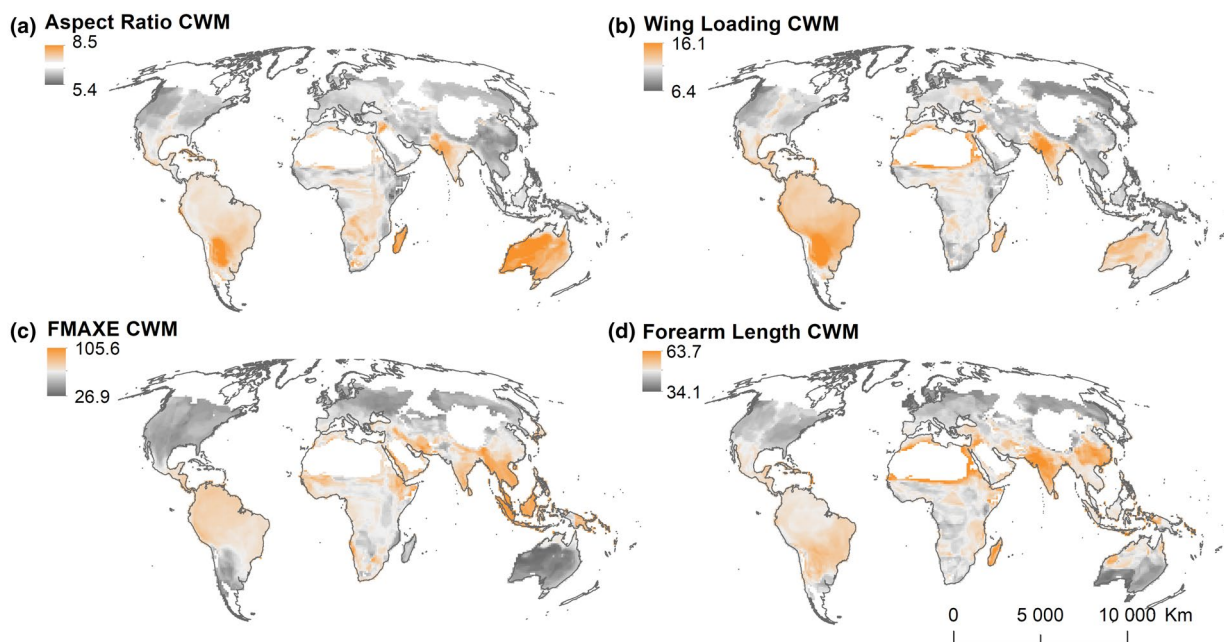


FIGURE 2 Geographical distributions of community weighted means (CWMs) for (a) aspect ratio, (b) wing loading, (c) FMAXE and (d) forearm length across global assemblages (median across 2,500 imputed dataset). FMAXE = frequency of maximum energy. The map is in equal-area Mollweide projection. [Colour figure can be viewed at wileyonlinelibrary.com]

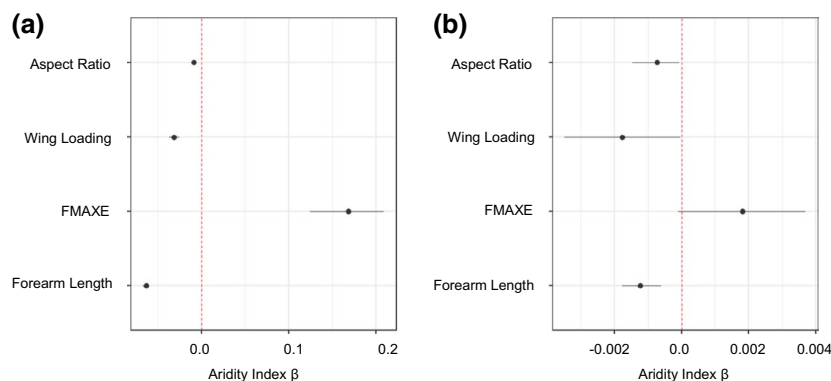


FIGURE 3 Summary of the results of (a) the assemblage-level analysis and (b) the species-level analysis. (a) Distribution (median and 95% confidence intervals) of the coefficients for the aridity index (AI) across the 2,500 models per trait. (b) Distribution (median and 95% confidence intervals) of the coefficients for the 5th percentile of aridity estimated across the 2,500 models per trait. Lower values of the AI correspond to higher aridity levels, and therefore for example, a negative coefficient indicates an increase in trait values with increasing aridity. The detected signal is considered highly probable if the 95% confidence intervals do not include zero, indicating that effects were consistent across models. FMAXE = frequency of maximum energy [Colour figure can be viewed at wileyonlinelibrary.com]

datasets, indicating the results are not affected by imputation uncertainty (Supporting Information Figures S4.11 and S4.12).

3.3 | Species-level analysis

Results from the PGLS models for the species-level analysis were largely consistent with the patterns revealed via the GAM approach for the assemblage-level analysis (Figure 3). Forearm length showed an increase with increasing aridity [across 2,500 models for each trait, median β_{aridity} (95% CIs) = -0.0012 (-0.0018 – -0.0006), $\text{se}_{\text{aridity}} = 0.0006 \pm 0$], and the same pattern was noticed for aspect ratio [-0.0007 (-0.0015 – -0.0001), $\text{se}_{\text{aridity}} = 0.0004 \pm 0$] and wing loading [-0.0018 (-0.0035 – 0), $\text{se}_{\text{aridity}} = 0.0011 \pm 0$], while FMAXE decreased [0.0018 (-0.0001 – 0.0037), $\text{se}_{\text{aridity}} = 0.0012 \pm 0$; Figure 3b and Supporting Information Figure S4.11]. Distribution of coefficients across all models produced indicated aridity as a correlate of forearm length, aspect ratio and wing loading, while the trend for FMAXE was less important (Figure 3b). Estimated Pagel's λ indicated high phylogenetic signal for forearm length and moderate for all other traits, and showed little variation across model replicates (forearm length: $\lambda = .82 \pm .02$, aspect ratio: $\lambda = .45 \pm .04$, wing loading: $\lambda = .59 \pm .04$, FMAXE: $\lambda = .58 \pm .04$).

4 | DISCUSSION

In this study, we provide novel insights into the functional responses of bats to aridity. In accordance with our hypotheses, we observed that increased aridity is associated with wing morphology and echolocation attributes suitable to move and forage in open spaces (higher aspect ratio and wing loading and lower FMAXE values) and with larger body sizes (greater forearm length). Despite the moderate levels of phylogenetic conservatism for the traits considered, these

patterns were consistent between the two different methodological approaches, indicating that our results are valid independently from the phylogenetic relationships among species.

4.1 | Patterns of wing and echolocation traits along the aridity gradient

As aridity increases, assemblages are increasingly characterized by long narrow wings and low echolocation frequencies. These trends reflect the changes in vegetation structure associated with aridity and likely result from environmental filters favouring species better adapted to structurally more open habitats (Salinas-Ramos et al., 2020). The prevalence of these features in deserts and semi-deserts is likely linked to the relaxation of the environmental pressures typically associated with flight in highly cluttered spaces. Our results suggest that, as openness of the habitat increases, manoeuvrability requirements to avoid background obstacles are reduced, while speed and cost-efficiency of flight to cover longer distances are favoured. Accordingly, there is an increased need to receive information about prey and physical obstacles further away in space to allow turning, and this is made possible by the use of lower echolocation frequencies (Denzinger & Schnitzler, 2013). These trends were confirmed by the species-level approach. However, the trend for FMAXE received less support, potentially due to the complex and plastic nature of echolocation that is poorly represented by a single parameter (Denzinger & Schnitzler, 2013). As arid lands are expected to expand under climate change (Huang et al., 2016), the trends observed in this study might gain further importance in the future. In fact, previous studies on bat functional traits have associated low values of wing aspect ratio with a higher extinction risk (Jones et al., 2003), likely due to the limited dispersal ability and smaller range sizes that are associated with low values for this trait (Jones et al., 2003; Luo et al., 2019, but

see also Varzinczak et al., 2020). The expansion of arid lands could result in more open habitats and, according to our findings, this could create less suitable conditions for clutter foragers and thus potentially increase their extinction risk.

In addition to a greater suitability to open spaces, greater mobility can also be advantageous for other aspects of bat ecology in arid environments. Accessing scattered water bodies and ephemeral and patchy food sources, conditions typical of desert and semi-desert habitats, likely leads to increased daily movements, as shown for example, for the desert-adapted species *Rhinopoma microphyllum* (Bell, 1980; Conenna et al., 2019; Egert-Berg et al., 2018; Korine & Pinshow, 2004). Higher wing loading and aspect ratio, as observed in our study for bat assemblages in more arid areas, allow for fast and energy-efficient flights, representing an advantage when facing long nightly commutes (Norberg & Rayner, 1987). Moreover, previous studies that have simulated the reduction in sizes of open water bodies, potentially expected with climate change, have shown that at local scales highly mobile bats might be more severely affected by these changes than manoeuvrable species due to difficulties in accessing the water (Hall et al., 2016). Further knowledge is needed on how bat communities are structured in space in relation to water bodies and on distances covered by species to access these resources, as well as on how local phenomena relate with the broad scale ones detected in this study.

The shift towards higher wing loading and aspect ratio values with increasing aridity is particularly well exemplified at the interface between the Oriental, Oceanian and Australian biogeographical regions. Here, aspect ratio and wing loading values progressively increase from the humid Southeast Asia, to the north-west of Australia and eventually reach the highest values in the Australian deserts. A similar pattern was detected in South America, with Amazonian assemblages displaying lower values than assemblages in the adjacent arid zones. However, South America overall stands out for its particularly high values of wing loading. These can be linked to the fact that, besides hosting a number of widespread species of open space foragers retaining particularly high values of wing loading (e.g. *Eumops underwoodi*, *Cynomops planirostris*, *Promops nasutus*, family Molossidae), South America also features numerous species belonging to the subfamilies of Stenodermatinae and Glossophaginae (family Phyllostomidae). These species exhibit exceptionally high wing loading despite being narrow-space foragers (Norberg & Rayner, 1987). All the regions mentioned above also hold corresponding trends in FMAXE, with progressively lower values as aridity increases, supporting the idea of these assemblages being dominated by open space species.

However, our results represent a simplification of complex patterns, and traits might respond to thresholds in aridity in a nonlinear manner, or diverge from main trends under extreme conditions also due to low species richness. For example, we also identify arid areas that display a prevalence of clutter-type values across both wing and echolocation traits, for example, the Arabian Peninsula (Figure 2). It is possible that, at extremely low environmental productivity, prey is occasionally more tightly associated with vegetation

patches, favouring species capable of foraging closer to the vegetation or gleaning prey from plants and the ground. Indeed, several desert-adapted species display such hunting strategies (e.g. *Plecotus christii*, *Asellia tridens*, *Anthrozous pallidus*; Bell, 1982; Korine & Pinshow, 2004). Further research is needed to explain the prevalence of narrow space foragers in some of the highest-aridity areas.

4.2 | Patterns of body size along the aridity gradient

The body size in bat assemblages increased with aridity. This pattern was strongly supported by the phylogenetic-controlled approach, indicating that species larger than expected by their phylogenetic position were more frequent in drier environments. Our results suggest that, given the overall small sizes and susceptibility of bats to cutaneous evaporative water loss, it might be advantageous to retain a large body size due to its relative lower surface-to-volume ratio. This is in accordance with James (1970), who stated that, when considering the gradient from humid to dry, larger sizes are expected to be associated with drier conditions. This trend might gain more relevance when seen in conjunction with the increase, at higher aridity, in wing loading, signalling a decrease in wing surface relative to body size. Similarly, Olalla-Tárraga et al. (2009) showed that anuran body size positively correlated with water deficit in the Neotropical Cerrado biome, and labelled this pattern as responding to the 'water availability hypothesis', according to which anurans from drier areas exhibit larger body sizes to reduce water loss and prevent desiccation. These patterns have also been previously associated with the 'water conservation hypothesis', which states that species are expected to be larger in more arid environments compared to mesic ones due to the advantage conferred by a larger body size in reducing desiccation rates (Nevo, 1973). While the relationship between bat body size and aridity has received limited attention in bats, the relationship with temperature has been extensively investigated, with various outcomes either confirming, refuting or contrasting Bergmann's rule (e.g. intraspecific: Jiang et al., 2019; Penone et al., 2018; interspecific: Safi et al., 2013). The divergence from Bergmann's rule in endotherms has been associated with size-independent adaptations to cold or warm climates (Medina et al., 2007), such as the ability to hibernate in bats. However, an interesting future avenue would be investigating whether these various identified trends between body size and temperature arise because of the combination of warm temperatures with various aridity levels, thus determining different outcomes depending on whether arid conditions are associated with either cold or warm temperatures (James, 1970).

4.3 | Limitations of the study

The results of this study are consistent across the different methodological approaches employed. Nevertheless, we would like to draw the reader's attention towards potential shortcomings of the study. First, the effects of aridity on bat traits detected across the analyses

are relatively weak. Many other factors, including other environmental variables, historical and local components are likely to combine to determine the trait values observed. Second, Hawkins et al. (2017) have recently highlighted that the repeated co-occurrence of species across grid cells, as is the case for the assemblage-level analysis in this study, can potentially increase the risk of Type I errors. While we encourage the reader to consider these caveats when evaluating the results of this study, we believe the validity of the study is not hindered since our aim was that of detecting overall effects of aridity on traits, rather than predicting trait values. Additionally, the results of the species-level analysis, which is insensitive to the bias caused by repeated species co-occurrences, support the overall robustness of conclusions.

5 | CONCLUSIONS

Taken together, our analyses revealed patterns of variation along the global aridity gradient in several important bat traits. Importantly, as aridity increases, we observed a prevalence of traits favourable to hunting in open spaces, as well as larger body sizes. These features might be advantageous in dry climates, allowing bats to exploit habitats with open structure while coping with scattered resources, as well as reduce evaporative water loss.

Environmental aridity poses a multitude of challenges to species and can be a driver of changes in local assemblage composition. Describing variation in functional traits along the aridity gradient, as presented in this study, can help us identify these changes and suggest how different groups of species might respond to the expected expansion in arid lands.

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AUTHOR CONTRIBUTIONS

IC, LS and DR conceived the ideas and designed methodology; IC collected the data; IC and LS analysed the data and MC, RR and AM provided important input into data interpretation. IC led the writing of the manuscript. All authors contributed critically to the drafts and gave final approval for publication.

DATA AVAILABILITY STATEMENT

The datasets supporting this article are available via the open-access repository Dryad (<https://doi.org/10.5061/dryad.tmpg4f4xm>).

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REFERENCES

- Aldridge, H. D. J. N., & Rautenbach, I. L. (1987). Morphology, echolocation and resource partitioning in insectivorous bats. *Journal of Animal Ecology*, 56, 763–778. <https://doi.org/10.2307/4947>
- Barnagaud, J. Y., Kissling, W. D., Tsiogiannis, C., Fisikopoulos, V., Vileger, S., Sekercioglu, C. H., & Svenning, J. C. (2017). Biogeographical, environmental and anthropogenic determinants of global patterns in bird taxonomic and trait turnover. *Global Ecology and Biogeography*, 26, 1190–1200. <https://doi.org/10.1111/geb.12629>
- Bell, G. P. (1980). Habitat use and response to patches of prey by desert insectivorous bats. *Canadian Journal of Zoology-Revue Canadienne De Zoologie*, 58, 1876–1883. <https://doi.org/10.1139/z80-256>
- Bell, G. P. (1982). Behavioral and ecological aspects of gleaning by a desert insectivorous bat, *Antrozous pallidus* (Chiroptera, Vespertilionidae). *Behavioral Ecology and Sociobiology*, 10, 217–223. <https://doi.org/10.1007/BF00299688>
- Bergmann, C. (1847). *Über die Verhältnisse der Wärmeökonomie der Thiere zu ihrer Grösse*. Göttinger Studien.
- Blackburn, T. M., & Gaston, K. J. (1996). Spatial patterns in the body sizes of bird species in the New World. *Oikos*, 77, 436–446. <https://doi.org/10.2307/3545933>
- Bondarenko, A., Kortner, G., & Geiser, F. (2014). Hot bats: Extreme thermal tolerance in a desert heat wave. *Naturwissenschaften*, 101, 679–685. <https://doi.org/10.1007/s00114-014-1202-2>
- Brum, F. T., Graham, C. H., Costa, G. C., Hedges, S. B., Penone, C., Radeloff, V. C., Rondinini, C., Loyola, R., & Davidson, A. D. (2017). Global priorities for conservation across multiple dimensions of mammalian diversity. *Proceedings of the National Academy of Sciences USA*, 114, 7641–7646. <https://doi.org/10.1073/pnas.1706461114>
- Buckley, L. B., & Jetz, W. (2008). Linking global turnover of species and environments. *Proceedings of the National Academy of Sciences USA*, 105, 17836–17841. <https://doi.org/10.1073/pnas.0803524105>
- Carpenter, R. E. (1969). Structure and function of the kidney and the water balance of desert bats. *Physiological Zoology*, 42, 288–302. <https://doi.org/10.1086/physzool.42.3.30155492>
- Carpenter, R. E. (1986). Flight physiology of intermediate-sized fruit bats (Pteropodidae). *Journal of Experimental Biology*, 120, 79–103.
- Collen, A. (2012). *The evolution of echolocation in bats: A comparative approach* (Doctoral dissertation). University College London.
- Conenna, I., López-Baucells, A., Rocha, R., Ripperger, S., & Cabeza, M. (2019). Movement seasonality in a desert-dwelling bat revealed by miniature GPS loggers. *Movement Ecology*, 7, 27. <https://doi.org/10.1186/s40462-019-0170-8>
- Cooke, R. S. C., Eigenbrod, F., & Bates, A. E. (2019). Projected losses of global mammal and bird ecological strategies. *Nature Communications*, 10, 2279. <https://doi.org/10.1038/s41467-019-10284-z>
- de Bello, F., Berg, M. P., Dias, A. T. C., Diniz-Filho, J. A. F., Götzenberger, L., Hortal, J., Ladle, R. J., & Lepš, J. (2015). On the need for phylogenetic 'corrections' in functional trait-based approaches. *Folia Geobotanica*, 50, 349–357. <https://doi.org/10.1007/s12224-015-9228-6>
- Demirtas, H., Freels, S. A., & Yucel, R. M. (2008). Plausibility of multivariate normality assumption when multiply imputing non-Gaussian continuous outcomes: A simulation assessment. *Journal of Statistical Computation and Simulation*, 78, 69–84. <https://doi.org/10.1080/10629360600903866>
- Denzinger, A., & Schnitzler, H. U. (2013). Bat guilds, a concept to classify the highly diverse foraging and echolocation behaviors of microchiropteran bats. *Frontiers in Physiology*, 4, 164. <https://doi.org/10.3389/fphys.2013.00164>

- Egert-Berg, K., Hurme, E. R., Greif, S., Goldstein, A., Harten, L., Herrera M., L. G., Flores-Martínez, J. J., Valdés, A. T., Johnston, D. S., Eitan, O., Borissov, I., Shipley, J. R., Medellín, R. A., Wilkinson, G. S., Goerlitz, H. R., & Yovel, Y. (2018). Resource ephemerality drives social foraging in bats. *Current Biology*, 28(3667–3673), e3665. <https://doi.org/10.1016/j.cub.2018.09.064>
- Estrada-Villegas, S., Meyer, C. F. J., & Kalko, E. K. V. (2010). Effects of tropical forest fragmentation on aerial insectivorous bats in a land-bridge island system. *Biological Conservation*, 143, 597–608. <https://doi.org/10.1016/j.biocon.2009.11.009>
- Farneda, F. Z., Rocha, R., López-Baucells, A., Groenenberg, M., Silva, I., Palmeirim, J. M., Bobrowiec, P. E. D., & Meyer, C. F. J. (2015). Trait-related responses to habitat fragmentation in Amazonian bats. *Journal of Applied Ecology*, 52, 1381–1391. <https://doi.org/10.1111/1365-2664.12490>
- Faurby, S., Davis, M., Pedersen, R. O., Schowaneck, S. D., Antonelli, A., & Svenning, J. C. (2018). PHYLACINE 1.2: The phylogenetic atlas of mammal macroecology. *Ecology*, 99, 2626.
- Fick, S. E., & Hijmans, R. J. (2017). WorldClim 2: New 1km spatial resolution climate surfaces for global land areas. *International Journal of Climatology*, 37, 4302–4315. <https://doi.org/10.1002/joc.5086>
- Findley, J. S., Studier, E. H., & Wilson, D. E. (1972). Morphologic properties of bat wings. *Journal of Mammalogy*, 53, 429–444. <https://doi.org/10.2307/1379035>
- Fletcher, R., & Fortin, M. (2018). *Spatial ecology and conservation modeling*. Springer.
- Fuller, A., Hetem, R. S., Maloney, S. K., & Mitchell, D. (2014). Adaptation to heat and water shortage in large, arid-zone mammals. *Physiology*, 29, 159–167. <https://doi.org/10.1152/physiol.00049.2013>
- Hall, L. K., Lambert, C. T., Larsen, R. T., Knight, R. N., & McMillan, B. R. (2016). Will climate change leave some desert bat species thirstier than others? *Biological Conservation*, 201, 284–292. <https://doi.org/10.1016/j.biocon.2016.07.020>
- Hawkins, B. A., Leroy, B., Rodríguez, M. Á., Singer, A., Vilela, B., Villalobos, F., Wang, X., & Zelený, D. (2017). Structural bias in aggregated species-level variables driven by repeated species co-occurrences: A pervasive problem in community and assemblage data. *Journal of Biogeography*, 44, 1199–1211. <https://doi.org/10.1111/jbi.12953>
- Hayes, M. J. L. J. P., Kurta, A., & Tuttle, M. D. (2007). *Bats in forests: Conservation and management*. JHU Press.
- Holt, B. G., Costa, G. C., Penone, C., Lessard, J.-P., Brooks, T. M., Davidson, A. D., Blair Hedges, S., Radeloff, V. C., Rahbek, C., Rondinini, C., & Graham, C. H. (2018). Environmental variation is a major predictor of global trait turnover in mammals. *Journal of Biogeography*, 45, 225–237. <https://doi.org/10.1111/jbi.13091>
- Huang, J., Yu, H., Guan, X., Wang, G., & Guo, R. (2016). Accelerated dryland expansion under climate change. *Nature Climate Change*, 6, 166–171.
- IUCN. (2019). *The IUCN Red List of Threatened Species*. Version 2019-3. <https://www.iucnredlist.org>
- James, F. C. (1970). Geographic size variation in birds and its relationship to climate. *Ecology*, 51, 365–390. <https://doi.org/10.2307/1935374>
- Jiang, T., Wang, J., Wu, H., Csorba, G., Puechmaile, S. J., Benda, P., Boireau, J., Toffoli, R., Courtois, J. Y., Nyssen, P., & Colombo, R. (2019). The patterns and possible causes of global geographical variation in the body size of the greater horseshoe bat (*Rhinolophus ferrumequinum*). *Journal of Biogeography*, 46, 2363–2377.
- Jones, K. E., Bielby, J., Cardillo, M., Fritz, S. A., O'Dell, J., Orme, C. D. L., Safi, K., Sechrest, W., Boakes, E. H., Carbone, C., Connolly, C., Cutts, M. J., Foster, J. K., Grenyer, R., Habib, M., Plaster, C. A., Price, S. A., Rigby, E. A., Rist, J., ... Purvis, A. (2009). PanTHERIA: A species-level database of life history, ecology, and geography of extant and recently extinct mammals. *Ecology*, 90, 2648. <https://doi.org/10.1890/08-1494.1>
- Jones K. E., & Purvis A. (1997). An optimum body size for mammals? Comparative evidence from bats. *Functional Ecology*, 11(6), 751–756. <http://dx.doi.org/10.1046/j.1365-2435.1997.00149.x>
- Jones, K. E., Purvis, A., & Gittleman, J. L. (2003). Biological correlates of extinction risk in bats. *The American Naturalist*, 161, 601–614. <https://doi.org/10.1086/368289>
- Korine, C., & Pinshow, B. (2004). Guild structure, foraging space use, and distribution in a community of insectivorous bats in the Negev Desert. *Journal of Zoology*, 262, 187–196. <https://doi.org/10.1017/S0952836903004539>
- Lawing, A. M., Eronen, J. T., Blois, J. L., Graham, C. H., & Polly, P. D. (2017). Community functional trait composition at the continental scale: The effects of non-ecological processes. *Ecography*, 40(5), 651–663. <https://doi.org/10.1111/ecog.01986>
- Luo, B., Santana, S. E., Pang, Y., Wang, M., Xiao, Y., & Feng, J. (2019). Wing morphology predicts geographic range size in vespertilionid bats. *Scientific Reports*, 9, 4526. <https://doi.org/10.1038/s41598-019-41125-0>
- Madley-Dowd, P., Hughes, R., Tilling, K., & Heron, J. (2019). The proportion of missing data should not be used to guide decisions on multiple imputation. *Journal of Clinical Epidemiology*, 110, 63–73. <https://doi.org/10.1016/j.jclinepi.2019.02.016>
- Maestre, F. T., Eldridge, D. J., Soliveres, S., Kéfi, S., Delgado-Baquerizo, M., Bowker, M. A., García-Palacios, P., Gaitán, J., Gallardo, A., Lázaro, R., & Berdugo, M. (2016). Structure and functioning of dryland ecosystems in a changing world. *Annual Review of Ecology, Evolution, and Systematics*, 47, 215–237. <https://doi.org/10.1146/annurev-ecolsys-121415-032311>
- Medina, A., Marti, D. A., & Bidau, C. J. (2007). Subterranean rodents of the genus *Ctenomys* (Caviomorpha, Ctenomyidae) follow the converse to Bergmann's rule. *Journal of Biogeography*, 34, 1439–1454. <https://doi.org/10.1111/j.1365-2699.2007.01708.x>
- Meiri, S. (2011). Bergmann's rule—what's in a name? *Global Ecology and Biogeography*, 20, 203–207. <https://doi.org/10.1111/j.1466-8238.2010.00577.x>
- Minnaar, I. A., Bennett, N. C., Chimimba, C. T., & McKechnie, A. E. (2014). Partitioning of evaporative water loss into respiratory and cutaneous pathways in Wahlberg's epauletted fruit bats (*Epomophorus wahlbergi*). *Physiological and Biochemical Zoology*, 87, 475–485.
- Monadjem, A., Taylor, P. J., Cotterill, E., & Schoeman, M. C. (2010). *Bats of southern and central Africa*. Wits University Press.
- Munoz-Garcia, A., Larrain, P., Ben-Hamo, M., Cruz-Neto, A., Williams, J. B., Pinshow, B., & Korine, C. (2016). Metabolic rate, evaporative water loss and thermoregulatory state in four species of bats in the Negev desert. *Comparative Biochemistry and Physiology A-Molecular & Integrative Physiology*, 191, 156–165. <https://doi.org/10.1016/j.cbpa.2015.10.010>
- Nakagawa, S., & Freckleton, R. P. (2008). Missing inaction: The dangers of ignoring missing data. *Trends in Ecology and Evolution*, 23, 592–596. <https://doi.org/10.1016/j.tree.2008.06.014>
- Nevo, E. (1973). Adaptive variation in size of cricket frogs. *Ecology*, 54, 1271–1281. <https://doi.org/10.2307/1934189>
- Norberg, U. M., & Rayner, J. M. (1987). Ecological morphology and flight in bats (Mammalia; Chiroptera): Wing adaptations, flight performance, foraging strategy and echolocation. *Philosophical Transactions of the Royal Society of London Series B, Biological Sciences*, 316, 335–427.
- O'Connell, M. A., & Hallett, J. G. (2019). Community ecology of mammals: Deserts, islands, and anthropogenic impacts. *Journal of Mammalogy*, 100, 1019–1043. <https://doi.org/10.1093/jmammal/gyz010>
- Olalla-Tárraga, M. A., Bini, L. M., Diniz-Filho, J. A., & Rodríguez, M. Á. (2010). Cross-species and assemblage-based approaches to Bergmann's rule and the biogeography of body size in *Plethodon* salamanders of eastern North America. *Ecography*, 33, 362–368. <https://doi.org/10.1111/j.1600-0587.2010.06244.x>

- Olalla-Tárraga, M. A., Diniz, J. A. F., Bastos, R. P., & Rodríguez, M. A. (2009). Geographic body size gradients in tropical regions: Water deficit and anuran body size in the Brazilian Cerrado. *Ecography*, 32, 581–590. <https://doi.org/10.1111/j.1600-0587.2008.05632.x>
- Oliveira, B. F., Machac, A., Costa, G. C., Brooks, T. M., Davidson, A. D., Rondinini, C., & Graham, C. H. (2016). Species and functional diversity accumulate differently in mammals. *Global Ecology and Biogeography*, 25, 1119–1130. <https://doi.org/10.1111/geb.12471>
- Oliveira, B. F., & Scheffers, B. R. (2019). Vertical stratification influences global patterns of biodiversity. *Ecography*, 42, 249. <https://doi.org/10.1111/ecog.03636>
- Pagel, M. (1999). Inferring the historical patterns of biological evolution. *Nature*, 401, 877–884. <https://doi.org/10.1038/44766>
- Paradis, E., & Schliep, K. (2019). ape 5.0: An environment for modern phylogenetics and evolutionary analyses in R. *Bioinformatics*, 35, 526–528. <https://doi.org/10.1093/bioinformatics/bty633>
- Penone, C., Davidson, A. D., Shoemaker, K. T., Di Marco, M., Rondinini, C., Brooks, T. M., Young, B. E., Graham, C. H., & Costa, G. C. (2014). Imputation of missing data in life-history trait datasets: Which approach performs the best? *Methods in Ecology and Evolution*, 5, 961–970. <https://doi.org/10.1111/2041-210X.12232>
- Penone, C., Kerbiriou, C., Julien, J.-F., Marmet, J., & Le Viol, I. (2018). Body size information in large-scale acoustic bat databases. *PeerJ*, 6, e5370. <https://doi.org/10.7717/peerj.5370>
- Pinheiro, J., Bates, D., DebRoy, S., Sarkar, D., & R Core Team. (2019). *nlme: Linear and nonlinear mixed effects models*. R package version 3.1-140. <https://CRAN.R-project.org/package=nlme>
- Rapacciuolo, G., Marín, J., Costa, G. C., Helmus, M. R., Behm, J. E., Brooks, T. M., Hedges, S. B., Radeloff, V. C., Young, B. E., & Graham, C. H. (2017). The signature of human pressure history on the biogeography of body mass in tetrapods. *Global Ecology and Biogeography*, 26, 1022–1034. <https://doi.org/10.1111/geb.12612>
- R Core Team (2019). *R: A language and environment for statistical computing*. R Foundation for Statistical Computing.
- Rodríguez, M. A., Olalla-Tárraga, M. A., & Hawkins, B. A. (2008). Bergmann's rule and the geography of mammal body size in the Western Hemisphere. *Global Ecology and Biogeography*, 17, 274–283. <https://doi.org/10.1111/j.1466-8238.2007.00363.x>
- Rymer, T. L., Pillay, N., & Schradin, C. (2016). Resilience to droughts in mammals: A conceptual framework for estimating vulnerability of a single species. *Quarterly Review of Biology*, 91, 133–176. <https://doi.org/10.1086/686810>
- Safi, K., Cianciaruso, M. V., Loyola, R. D., Brito, D., Armour-Marshall, K., & Diniz-Filho, J. A. F. (2011). Understanding global patterns of mammalian functional and phylogenetic diversity. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 366, 2536–2544. <https://doi.org/10.1098/rstb.2011.0024>
- Safi, K., Meiri, S., & Jones, K. E. (2013). Evolution of body size in bats. In F. A. Smith & S. K. Lyons (Eds.), *Animal body size: Linking pattern and process across space, time, and taxonomic group*. (95–151). University of Chicago Press.
- Safriel, U., Adeel, Z., Niemeijer, D., Puigdefabregas, J., White, R., Lal, R., & McNab, D. (2005). Dryland systems. In R. Hassan R. Scholes & N. Ash (Ed.), *Ecosystems and human well-being: Current state and trends* (pp. 623–662). Island Press.
- Santini, L., González-Suárez, M., Rondinini, C., & Di Marco, M. (2017). Shifting baseline in macroecology? Unravelling the influence of human impact on mammalian body mass. *Diversity and Distributions*, 23, 640–649. <https://doi.org/10.1111/ddi.12555>
- Schwimmer, H., & Haim, A. (2009). Physiological adaptations of small mammals to desert ecosystems. *Integrative Zoology*, 4, 357–366. <https://doi.org/10.1111/j.1749-4877.2009.00176.x>
- Simmons, N. (2003). Evolution of ecological diversity in bats. In T. Kunz, & M. B. Fenton (Eds.), *Bat ecology* (pp. 493–534). University of Chicago Press.
- Torres-Romero, E. J., Morales-Castilla, I., & Olalla-Tárraga, M. Á. (2016). Bergmann's rule in the oceans? Temperature strongly correlates with global interspecific patterns of body size in marine mammals. *Global Ecology and Biogeography*, 25, 1206–1215. <https://doi.org/10.1111/geb.12476>
- Trabucco, A., & Zomer, R. (2019). *Global aridity index and potential evapotranspiration (ET0) climate database v2*. https://figshare.com/articles/dataset/Global_Aridity_Index_and_Potential_Evapotranspiration_ET0_Climate_Database_v2/7504448/3
- Upham, N. S., Esselstyn, J. A., & Jetz, W. (2019). Inferring the mammal tree: Species-level sets of phylogenies for questions in ecology, evolution, and conservation. *PLoS Biology*, 17(12), e3000494. <https://doi.org/10.1371/journal.pbio.3000494>
- Van Buuren, S. (2018). *Flexible imputation of missing data*. CRC Press.
- Van Buuren, S., & Groothuis-Oudshoorn, K. (2011). mice: Multivariate imputation by chained equations in R. *Journal of Statistical Software*, 45(3), 1–68.
- Varzinczak, L. H., Moura, M. O., & Passos, F. C. (2020). Strong but opposing effects of climatic niche breadth and dispersal ability shape bat geographical range sizes across phylogenetic scales. *Global Ecology and Biogeography*, 29, 1929–1939. <https://doi.org/10.1111/geb.13163>
- Violle, C., Reich, P. B., Pacala, S. W., Enquist, B. J., & Kattge, J. (2014). The emergence and promise of functional biogeography. *Proceedings of the National Academy of Sciences USA*, 111, 13690–13696. <https://doi.org/10.1073/pnas.1415442111>
- Wickham, H. (2011). The split-apply-combine strategy for data analysis. *Journal of Statistical Software*, 40, 1–29. <https://doi.org/10.18637/jss.v040.i01>
- Williams, J. B., & Tieleman, B. I. (2005). Physiological adaptation in desert birds. *BioScience*, 55(5), 416–425.
- Wilman, H., Belmaker, J., Simpson, J., De La Rosa, C., Rivadeneira, M. M., & Jetz, W. (2014). EltonTraits 1.0: Species-level foraging attributes of the world's birds and mammals. *Ecology*, 95, 2027. <https://doi.org/10.1890/13-1917.1>
- Wood, S. N. (2011). Fast stable restricted maximum likelihood and marginal likelihood estimation of semiparametric generalized linear models. *Journal of the Royal Statistical Society: Series B (Statistical Methodology)*, 73, 3–36. <https://doi.org/10.1111/j.1467-9868.2010.00749.x>
- Wood, S. N. (2017). *Generalized additive models: An introduction with R*. CRC Press.
- Zambrano-Bigiarini, M. (2017). *Goodness-of-fit functions for comparison of simulated and observed hydrological time series*. R package version 0.3-10. <http://hzambran.github.io/hydroGOF/>

BIOSKETCH

Irene Conenna is an ecologist interested in the different behavioural and functional responses of bats to aridity, as well as in the implications under global changes and conservation issues at large.

SUPPORTING INFORMATION

Additional Supporting Information may be found online in the Supporting Information section.

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APPENDIX A

DATA SOURCES

- Andriafidison, D., Kofoky, A., Mbohoahy, T., Racey, P. A., & Jenkins, R. K. (2007). Diet, reproduction and roosting habits of the Madagascar free-tailed bat, *Otomops madagascariensis* Dorst, 1953 (Chiroptera: Molossidae). *Acta Chiropterologica*, **9**, 445–450.
- Arias-Aguilar, A. P. (2017). *Morcegos insetívoros aéreos neotropicais: identificação acústica e padrões de estruturação de assembleias*. (Master's dissertation), Universidade Federal do Rio Grande do Sul, Porto Alegre.
- Arias-Aguilar, A., Hintze, F., Aguiar, L. M., Rufray, V., Bernard, E., & Pereira, M. J. R. (2018). Who's calling? Acoustic identification of Brazilian bats. *Mammal Research*, **63**, 231–253.
- Armstrong, K. N., & Aplin, K. P. (2009). Bats of the Muller Range, Papua New Guinea. In S. J., Richards, B. G., Gamui (Eds.) *Rapid Biological Assessments of the Nakanai Mountains and the upper Strickland Basin: surveying the biodiversity of Papua New Guinea's sub-lime karst environments* (pp. 222–234). BioOne.
- Arroyo-Cabrales, J., Kalko, E. K. V., LaVal, R. K., Maldonado, J. E., Medellín, R. A., Polaco, O. J., & Rodríguez-Herrera, B. (2005). Rediscovery of the Mexican flat-headed bat *Myotis planiceps* (Vespertilionidae). *Acta Chiropterologica*, **7**, 309–314.
- Baker, R. J., McDonough, M. M., Swier, V. J., Larsen, P. A., Carrera, J. P., & Ammerman, L. K. (2009). New species of bonneted bat, genus *Eumops* (Chiroptera: Molossidae) from the lowlands of western Ecuador and Peru. *Acta Chiropterologica*, **11**, 1–13.
- Barataud, M., Giosa, S., Leblanc, F., Rufray, V., Disca, T., Tillon, L., Delaval, M., Haquart, A., & Dewynter, M. (2013). Identification et écologie acoustique des chiroptères de Guyane française. *Le Rhinophe*, **19**, 103–145.
- Bates, P. J. J., Rossiter, S. J., Suyanto, A., & Kingston, T. (2007). A new species of *Hipposideros* (Chiroptera: Hipposideridae) from Sulawesi. *Acta Chiropterologica*, **9**, 13–26.
- Beguelini, M. R., Puga, C. C., Taboga, S. R., & Mориelle-Versute, E. (2013). Annual reproductive cycle of males of the flat-faced fruit-eating bat, *Artibeus planirostris* (Chiroptera: Phyllostomidae). *General and Comparative Endocrinology*, **185**, 80–89.
- Benda, P., Gazaryan, S., & Vallo, P. (2016). On the distribution and taxonomy of bats of the *Myotis mystacinus* morphogroup from the Caucasus region (Chiroptera: Vespertilionidae). *Turkish Journal of Zoology*, **40**, 842–863.
- Benda, P., Georgiakakis, P., Dietz, C., Hanák, V., Galanaki, K., Markantonatou, V., Chudáková, A., Hulva, P., & Horáček, I. (2008). Bats (Mammalia: Chiroptera) of the Eastern Mediterranean and Middle East. Part 7. The bat fauna of Crete, Greece. *Acta Societatis Zoologicae Bohemicae*, **72**, 105–190.
- Benda, P., & Tsytsulina, K. (2000). Taxonomic revision of *Myotis mystacinus* group (Mammalia: Chiroptera). *Acta Societatis Zoologicae Bohemicae*, **64**, 331–398.
- Bernard, E. (2001). Vertical stratification of bat communities in primary forests of Central Amazon, Brazil. *Journal of Tropical Ecology*, **17**, 115–126.
- Beuneux, G. (2004). Morphometrics and ecology of *Myotis cf. puniceus* (Chiroptera, Vespertilionidae) in Corsica. *Mammalia*, **68**, 269–273.
- Briones-Salas, M., Peralta-Pérez, M., & García-Luis, M. (2013). Acoustic characterization of new species of bats for the State of Oaxaca, Mexico. *Therya*, **4**, 15–32.
- Calderón, W., & Pacheco, V. (2012). First report of *Artibeus bogotensis* Andersen, 1906 (Chiroptera: Phyllostomidae) for Peru. *Check List*, **8**, 1333–1336.
- Churchill, S., Hall, L., & Helman, P. (1984). Observations on long-eared bats (Vespertilionidae: Nyctophilus) from Northern Australia. *Australian Mammalogy*, **7**, 17–28.
- Collen, A. (2012). *The evolution of echolocation in bats: A comparative approach* (Doctoral dissertation). University College, London.
- Colombo, G. T. (2017). *Influência dos traços funcionais na distribuição de morcegos insetívoros aéreos em uma paisagem insularizada* (Master's dissertation). Instituto Nacional de Pesquisas da Amazônia.
- Corcoran, A. J. (2007). *Automated acoustic identification of nine bat species of the eastern United States* (Master's dissertation). Humboldt State University, Arcata.
- Crome, F. H. J., & Richards, G. C. (1988). Bats and gaps—microchiropteran community structure in a Queensland rain-forest. *Ecology*, **69**, 1960–1969.
- Csorba, G., & Bates, P. J. J. (2005). Description of a new species of *Murina* from Cambodia (Chiroptera: Vespertilionidae: Murinae). *Acta Chiropterologica*, **7**, 1–7.
- Csorba, G., & Lee, L. L. (1999). A new species of vespertilionid bat from Taiwan and a revision of the taxonomic status of *Arielulus* and *Thainycteris* (Chiroptera: Vespertilionidae). *Journal of Zoology*, **248**, 361–367.
- Dalhouri, R., Aissa, P., & Aulagnier, S. (2016). Seasonal variations of sexual size dimorphism in two Mediterranean bat species from Tunisia: The Kuhl's pipistrelle (*Pipistrellus kuhlii*) and the Isabelline serotine (*Eptesicus isabellinus*). *Folia Zoologica*, **65**(2), 157–163. <https://doi.org/10.25225/fozo.v65.i2.a11.2016>
- Davalos, L. M. (2004). A new choacoan species of *Lonchophylla* (Chiroptera : Phyllostomidae). *American Museum Novitates*, **2004**(3426), 1–14.
- Davie, H. S., Murdoch, J. D., Naran, N.-E., Ariunbold, J., Batdorj, S., & Reading, R. P. (2012). Bat diversity at Ikh Nart nature reserve, Mongolia. *Mongolian Journal of Biological Sciences*, **10**, 33–40.
- Davis, W. B. (1970). A review of the small fruit bats (genus *Artibeus*) of Middle America: Part II. *The Southwestern Naturalist*, **14**, 389–402.
- Decher, J., & Fahr, J. (2005). *Hipposideros cyclops*. *Mammalian Species*, **2005**(763), 1–7. <https://doi.org/10.2307/3504339>
- Delaval, M., Henry, M., & Charles-Dominique, P. (2005). Interspecific competition and niche partitioning: Example of a neotropical rainforest bat community. *Revue d'Ecologie-La Terre Et La Vie*, **60**, 149–165.
- Dias, D., de Carvalho, W. D., Teixeira, T. S. M., Tavares, D., da Silva Xavier, B., do Valle, E. L. V., & Esbérard, C. E. L. (2015). First record of *Myotis izecksohni* (Chiroptera, Vespertilionidae) for the Atlantic Forest of Minas Gerais, southeastern Brazil. *Mastozoologia neotropical*, **22**, 149–153.

- Douangboubpha, B., Bumrungsri, S., Satasook, C., Wanna, W., Soisook, P., & Bates, P. J. J. (2016). Morphology, genetics and echolocation calls of the genus *Kerivoula* (Chiroptera: Vespertilionidae: Kerivoulinae) in Thailand. *Mammalia*, **80**, 21–47.
- Emrich, M. A., Clare, E. L., Symondson, W. O., Koenig, S. E., & Fenton, M. B. (2014). Resource partitioning by insectivorous bats in Jamaica. *Molecular Ecology*, **23**, 3648–3656.
- Faurby, S., Davis, M., Pedersen, R. O., Schowaneck, S. D., Antonelli, A., & Svenning, J. C. (2018). PHYLACINE 1.2: The phylogenetic atlas of mammal macroecology. *Ecology*, **99**, 2626–2626.
- Fazzolari-Corrêa, S. (1994). *Lasiurus ebenus*, a new vespertilionid bat from southeastern Brasil. *Mammalia*, **58**, 119–124.
- Fenton, M. B., & Bell, G. P. (1981). Recognition of species of insectivorous bats by their echolocation calls. *Journal of Mammalogy*, **62**, 233–243.
- Fenton, M. B., Whitaker, J. O. Jr, Vonhof, M. J., Waterman, J. M., Pedro, W. A., Aguiar, L., Baumgarten, J. E., Bouchard, S., Faria, D. M., Portfors, C. V., & Rautenbach, N. I. (1999). The diet of bats from Southeastern Brazil: The relation to echolocation and foraging behaviour. *Revista Brasileira de Zoologia*, **16**, 1081–1085.
- Foley, N. M., Goodman, S. M., Whelan, C. V., Puechmaile, S. J., & Teeling, E. (2017). Towards navigating the Minotaur's labyrinth: Cryptic diversity and taxonomic revision within the speciose genus *Hipposideros* (Hipposideridae). *Acta Chiropterologica*, **19**, 1–18.
- Frank, H. K., Frishkoff, L. O., Mendenhall, C. D., Daily, G. C., & Hadly, E. A. (2017). Phylogeny, traits, and biodiversity of a neotropical bat assemblage: Close relatives show similar responses to local deforestation. *The American Naturalist*, **190**, 200–212.
- Fukui, D., Hill, D. A., Sun-Sook, K., & Han, S.-H. (2015). Echolocation call structure of fourteen bat species in Korea. *Animal Systematics, Evolution & Diversity*, **31**, 160–175.
- Fukui, D., Mochida, M., Yamamoto, A., & Kawai, K. (2013). Roost and echolocation call structure of the Alashanian pipistrelle *Hypsugo alaschanicus*: First confirmation as a resident species in Japan. *Mammal Study*, **38**, 61–66.
- Fullard, J. H., Koehler, C., Surlykke, A., & McKenzie, N. L. (1991). Echolocation ecology and flight morphology of insectivorous bats (Chiroptera) in South-Western Australia. *Australian Journal of Zoology*, **39**, 427–438.
- Funakoshi, K., & Kunisaki, T. (2000). On the validity of *Tadarida latouchei*, with reference to morphological divergence among *T. latouchei*, *T. insignis* and *T. teniotis* (Chiroptera, Molossidae). *Mammal Study*, **25**, 115–123.
- Furey, N. M., & Racey, P. A. (2016). Can wing morphology inform conservation priorities for Southeast Asian cave bats? *Biotropica*, **48**, 545–556.
- Genoways, H. H., & Baker, R. J. (1996). A new species of the genus *Rhogeessa*, with comments on geographic distribution and speciation in the genus. *Mammalogy Papers: University of Nebraska State Museum*, 248.
- Glass, B. P., & Ward, C. M. (1959). Bats of the genus *Myotis* from Oklahoma. *Journal of Mammalogy*, **40**, 194–201.
- Gonzalez-Ruiz, N., Ramirez-Pulido, J., & Arroyo-Cabral, J. (2011). A new species of mastiff bat (Chiroptera: Molossidae: Molossus) from Mexico. *Mammalian Biology*, **76**, 461–469.
- Goodman, S. M., & Cardiff, S. G. (2004). A new species of *Chaerephon* (Molossidae) from Madagascar with notes on other members of the family. *Acta Chiropterologica*, **6**, 227–248.
- Goodwin, G. G. (1958). *Bats of the genus Rhogeessa*. American Museum of Natural History.
- Goudy-Trainor, A., & Freeman, P. W. (2002). Call parameters and facial features in bats: A surprising failure of form following function. *Acta Chiropterologica*, **4**, 1–16.
- Gregorin, R., & Chiquito, E. A. (2010). Revalidation of *Promops davisoni* Thomas (Molossidae). *Chiroptera Neotropical*, **16**, 648–660.
- Gregorin, R., Tahara, A. S., & Buzzato, D. F. (2011). *Molossus aztecus* and other small Molossus (Chiroptera: Molossidae) in Brazil. *Acta Chiropterologica*, **13**, 311–317.
- Hackett, T. D., Holderied, M. W., & Korine, C. (2017). Echolocation call description of 15 species of Middle-Eastern desert dwelling insectivorous bats. *Bioacoustics-the International Journal of Animal Sound and Its Recording*, **26**, 217–235.
- Happold, M., & Happold, D. C. (2013). *Mammals of Africa: Hedgehogs, shrews and bats*. Bloomsbury Publishing.
- Haynes, M. A., & Lee, T. E. (2004). *Artibeus obscurus*. *Mammalian Species*, **2004**(752), 1–5.
- Heaney, L. R., Balete, D. S., Alviola, P., Rickart, E. A., & Ruedi, M. (2012). *Nyctalus plancyi* and *Falsistrellus petersi* (Chiroptera: Vespertilionidae) from northern Luzon, Philippines: Ecology, phylogeny, and biogeographic implications. *Acta Chiropterologica*, **14**, 265–278.
- Higginbotham, J. L., Ammerman, L. K., & Dixon, M. T. (1999). First record of *Lasiurus xanthinus* (Chiroptera : Vespertilionidae) in Texas. *The Southwestern Naturalist*, **44**, 343–347.
- Hill, J. (1964). Notes on a collection of bats from Figuig, Morocco. *Mammalia*, **28**, 83–87.
- Horta, P., Raposeira, H., Santos, H., Alves, P., Palmeirim, J., Godinho, R., Jones, G. and Rebelo, H., & Rebelo, H. (2015). Bats' echolocation call characteristics of cryptic Iberian *Eptesicus* species. *European Journal of Wildlife Research*, **61**, 813–818.
- Huang, J. C., Nur Izzati, A., Nurul-Ain, E., Shahrul, A. M. S., Lim, L. (2019). The first record of *Coelops frithii* (Blyth, 1848) to Penang, Malaysia, with a note of the potential acoustic plasticity of the species. *Journal of Bat Research and Conservation*, **12**, 21–26.
- Hughes, A. C., Satasook, C., Bates, P. J. J., Soisook, P., Sritongchuay, T., Jones, G., & Bumrungsri, S. (2011). Using echolocation calls to identify Thai bat species: Vespertilionidae, Emballonuridae, Nycteridae and Megadermatidae. *Acta Chiropterologica*, **13**, 447–455.
- Jacobs, D. S., Barclay, R. M., & Walker, M. H. (2007). The allometry of echolocation call frequencies of insectivorous bats: Why do some species deviate from the pattern? *Oecologia*, **152**, 583–594.
- Jacobs, D. S., Eick, G. N., Schoeman, M. C., & Matthee, C. A. (2006). Cryptic species in an insectivorous bat, *Scotophilus dinganii*. *Journal of Mammalogy*, **87**, 161–170.

- Jargalsaikhan, A. (2016). Bat study in the Kharaa region, Mongolia. *Journal of Asia-Pacific Biodiversity*, **9**, 107–115.
- Jennings, J. B., Best, T. L., Rainey, J. C., & Burnett, S. E. (2000). *Molossus pretiosus*. *Mammalian Species*, **2000**(635), 1–3.
- Jennings, N. V., Parsons, S., Barlow, K. E., & Gannon, M. R. (2004). Echolocation calls and wing morphology of bats from the West Indies. *Acta Chiropterologica*, **6**, 75–90.
- Jiang, T. L., You, Y. Y., Liu, S., Lu, G. J., Wang, L., Wu, H., ... Feng, J. (2013). Factors affecting geographic variation in echolocation calls of the endemic *Myotis davidii* in China. *Ethology*, **119**, 881–890.
- Jones, G., & Siemers, B. M. (2011). The communicative potential of bat echolocation pulses. *Journal of Comparative Physiology A*, **197**, 447–457.
- Jones, K. E., Bielby, J., Cardillo, M., Fritz, S. A., O'Dell, J., Orme, C. D. L., ... Carbone, C. (2009). PanTHERIA: A species-level database of life history, ecology, and geography of extant and recently extinct mammals. *Ecology*, **90**, 2648–2648.
- Jones, K. E., Purvis, A., & Gittleman, J. L. (2003). Biological correlates of extinction risk in bats. *The American Naturalist*, **161**, 601–614.
- Jung, K., & Threlfall, C. G. (2018). Trait-dependent tolerance of bats to urbanization: A global meta-analysis. *Proceedings of the Royal Society B: Biological Sciences*, **285**, 20181222.
- Kanuch, P., Aghova, T., Meheretu, Y., Sumbera, R., & Bryja, J. (2015). New discoveries on the ecology and echolocation of the heart-nosed bat *Cardioderma cor* with a contribution to the phylogeny of Megadermatidae. *African Zoology*, **50**, 53–57.
- Kingdon, J., Happold, D., Butynski, T., Hoffmann, M., Happold, M., & Kalina, J. (2013). *Mammals of Africa* (Vol. 1). A&C Black.
- Kingston, T., & Rossiter, S. J. (2004). Harmonic-hopping in Wallace's bats. *Nature*, **429**, 654–657.
- Korine, C., & Kalko, E. K. V. (2005). Fruit detection and discrimination by small fruit-eating bats (Phyllostomidae): Echolocation call design and olfaction. *Behavioral Ecology and Sociobiology*, **59**, 12–23.
- Kwiecinski, G. G., Pedersen, S. C., Genoways, H. H., Larsen, P. A., Larsen, R. J., Hoffman, J. D., ... Baker, R. J. (2018). Bats of St. Vincent, Lesser Antilles. *Special Publications / Texas Tech University Museum*, **68**.
- Leal, E. S. B., Silva, D. Q., Ramalho, D. d. F., Miller, B. G., Passos Filho, P. d. B., Neto, J. P., Queiróz-Guerra, D., Moura, G. J. B. D., Lyra-Neves, R. M. D., & Telino-Júnior, W. R. (2013). Extension of the geographical distribution of *Lonchophylla dekeyseri* Taddei, Vizotto and Sazima, 1983 (Chiroptera: Phyllostomidae): New record in north-eastern Brazil. *Chiroptera Neotropical*, **19**, 1220–1225.
- Lee, T. E. Jr, Burneo, S. F., Cochran, T. J., & Chavez, D. (2010). *Small mammals of Santa Rosa, southwestern Imbabura Province, Ecuador*. Occasional Papers, Museum of Texas Tech University.
- López-Baucells, A., Rocha, R., Tavares, V. D., Moras, L. M., Silva, S. E., Bobrowiec, P. E. D., & Meyer, C. F. J. (2018). Molecular, morphological and acoustic identification of *Eumops maurus* and *Eumops hansae* (Chiroptera: Molossidae) with new reports from Central Amazonia. *Tropical Zoology*, **31**, 1–20.
- López-Baucells, A., Rocha, R., Webala, P., Nair, A., Uusitalo, R., Sironen, T., & Forbes, K. M. (2016). Rapid assessment of bat diversity in the Taita Hills Afromontane cloud forests, southeastern Kenya. *Barbastella*, **9**, 1–10.
- Luo, B., Santana, S. E., Pang, Y., Wang, M., Xiao, Y., & Feng, J. (2019). Wing morphology predicts geographic range size in vespertilionid bats. *Scientific Reports*, **9**, 4526.
- Macswiney, M. C., Bolívar, B., Clarke, F. M., & Racey, P. A. (2006). Nuevos registros de *Pteronotus personatus* y *Cynomops mexicanus* (Chiroptera) en el estado de Yucatán, México. *Revista Mexicana de Mastozoología*, **10**, 80–87.
- Marin-Vasquez, A., Ramírez-Chaves, H. E., Rivas-Pava, P., & Mantilla-Meluk, H. (2014). New records of the Western rounded ear bat, *Lophostoma occidentale* (Davis & Carter, 1978) (Chiroptera: Phyllostomidae), from Colombia. *Check List*, **11**, 1513.
- Marinello, M. M., & Bernard, E. (2014). Wing morphology of Neotropical bats: A quantitative and qualitative analysis with implications for habitat use. *Canadian Journal of Zoology-Revue Canadienne De Zoologie*, **92**, 141–147.
- McCarthy, T. J., Albuja, L., & Alberico, M. S. (2006). A new species of Chocóan *Sturnira* (Chiroptera: Phyllostomidae: Stenodermatinae) from western Ecuador and Colombia. *Annals of Carnegie Museum*, **75**, 97–109.
- McKean, J. L., & Calaby, J. J. M. (1968). A new genus and two new species of bats from New Guinea. *Mammalia*, **32**, 372–378.
- Meyer, C. F. J., Frund, J., Lizano, W. P., & Kalko, E. K. V. (2008). Ecological correlates of vulnerability to fragmentation in Neotropical bats. *Journal of Applied Ecology*, **45**, 381–391.
- Miller, G. S., & Allen, G. M. (1928). *The American bats of the genera Myotis and Pizonyx* (Vol. 144). US Government Printing Office.
- Milne, D., Reardon, T., & Watt, F. (2003). New records for the Arnhem sheath-tail bat *Taphozous kapalgensis* (Chiroptera: Emballonuridae) from voucher specimens and Anabat recordings. *Australian Zoologist*, **32**, 439–445.
- Miranda, J. M. D., Bernardi, I. P., Sponchiado, J., & Passos, F. C. (2013). The taxonomic status of *Myotis levis levis* and *Myotis levis dinellii* (Mammalia: Chiroptera: Vespertilionidae). *Zoologia*, **30**(5), 513–518.
- Molinari, J. (1994). A new species of *Anoura* (Mammalia Chiroptera Phyllostomidae) from the Andes of northern South America. *Tropical Zoology*, **7**, 73–86.
- Monadjem, A., Conenna, I., Taylor, P. J., & Schoeman, M. C. (2018). Species richness patterns and functional traits of the bat fauna of arid southern Africa. *Hystrix-Italian Journal of Mammalogy*, **29**, 19–24.
- Monadjem, A., Schoeman, M. C., Reside, A., Pio, D. V., Stoffberg, S., Bayliss, J., Cotterill, F. P. D., Curran, M., Kopp, M., & Taylor, P. J. (2010). A recent inventory of the bats of Mozambique with documentation of seven new species for the country. *Acta Chiropterologica*, **12**, 371–391.
- Mora, E. C., Ibanez, C., Macias, S., Juste, J., Lopez, I., & Torres, L. (2011). Plasticity in the echolocation inventory of *Mormopterus minutus* (Chiroptera, Molossidae). *Acta Chiropterologica*, **13**(1), 179–187.
- Mora, J. M., Marineros, L., & Lopez, L. I. (2014). First record of the striped yellow-eared bat, *Vampyriscus nymphaea*, (Stenodermatinae, Phyllostomidae) in Honduras. *Caribbean Journal of Science*, **48**, 49–51.

- Moras, L. M., Gregorin, R., Sattler, T., & Tavares, V. D. (2018). Uncovering the diversity of dog-faced bats of the genus *Cynomops* (Chiroptera: Molossidae), with the redescription of *C. milleri* and the description of two new species. *Mammalian Biology*, **89**, 37–51.
- Muijres, F. T., Henningsson, P., Stuijver, M., & Hedenstrom, A. (2012). Aerodynamic flight performance in flap-gliding birds and bats. *Journal of Theoretical Biology*, **306**, 120–128.
- Murariu, D., & Gheorghiu, V. (2010). Şura Mare cave (Romania)-the most important known hibernating roost for *Pipistrellus pygmaeus* Leach, 1825 (Chiroptera: Vespertilionidae). *Travaux du Muséum National d'Histoire Naturelle "Grigore Antipa"*, **53**, 329–338.
- Nader, I. A. (1975). On the bats (Chiroptera) of the Kingdom of Saudi Arabia. *Journal of Zoology*, **176**, 331–340.
- Neuweiler, G., Metzner, W., Heilmann, U., Rubsamen, R., Eckrich, M., & Costa, H. H. (1987). Foraging behavior and echolocation in the rufous horseshoe bat (*Rhinolophus Rouxi*) of Sri-Lanka. *Behavioral Ecology and Sociobiology*, **20**, 53–67.
- Novaes, R. L. M., Laurindo, R. S., Dornas, R. A., Esbérard, C. E. L., & Bueno, C. (2018). On a collision course: The vulnerability of bats to roadkills in Brazil. *Mastozoología neotropical*, **25**, 115–128.
- O'Farrell, M. J., & Miller, B. W. (1999). Use of vocal signatures for the inventory of free-flying Neotropical bats. *Biotropica*, **31**, 507–516.
- Obrist, M. K., Fenton, M. B., Eger, J. L., & Schlegel, P. A. (1993). What ears do for bats—a comparative-study of pinna sound pressure transformation in Chiroptera. *Journal of Experimental Biology*, **180**, 119–152.
- Ossa, G., Bonacic, C., & Barquez, R. M. (2015). First record of *Histiotus laephotis* (Thomas, 1916) from Chile and new distributional information for *Histiotus montanus* (Phillipi and Landbeck, 1861) (Chiroptera, Vespertilionidae). *Mammalia*, **79**, 457–461.
- Ossa, G., Díaz, M. M., & Barquez, R. M. (2019). *Lasiurus varius* (Chiroptera: Vespertilionidae). *Mammalian Species*, **51**, 119–127.
- Ossa, G., Lilley, T. M., Ugarte-Núñez, J., Ruokolainen, L., Vilches, K., Valladares-Faúndez, P., & Yung, V. (2018). First record of *Promops davisoni* (Thomas, 1921) (Chiroptera, Molossidae) from Chile and a description of its echolocation calls. *Mastozoología neotropical*, **25**, 129–137.
- Pacheco, V., Cadenillas, R., Velazco, S., Salas, E., & Fajardo, U. (2007). Noteworthy bat records from the Pacific Tropical rainforest region and adjacent dry forest in northwestern Peru. *Acta Chiropterologica*, **9**, 409–422.
- Pacheco, V., Solari, S., & Velazco, P. M. (2004). A new species of *Carollia* (Chiroptera: Phyllostomidae) from the Andes of Peru and Bolivia (Vol. 236). Occasional Papers, Museum of Texas Tech University.
- Papadatou, E., Butlin, R. K., & Altringham, J. D. (2008). Identification of bat species in Greece from their echolocation calls. *Acta Chiropterologica*, **10**, 127–143.
- Phauk, S., Phen, S., & Furey, N. (2013). Cambodian bat echolocation: A first description of assemblage call parameters and assessment of their utility for species identification. *Cambodian Journal of Natural History*, **1**, 16–26.
- Pineda, W., Rodríguez-Herrera, B., & Timm, R. M. (2008). Rediscovery, ecology, and identification of rare free-tailed bats (Chiroptera: Molossidae) in Costa Rica. *Acta Chiropterologica*, **10**, 97–102.
- Pottie, S. A., Lane, D. J. W., Kingston, T., & Lee, B. P. Y. H. (2005). The microchiropteran bat fauna of Singapore. *Acta Chiropterologica*, **7**, 237–247.
- Pye, J. D. (1972). Bimodal distribution of constant frequencies in some hipposiderid bats (Mammalia-Hipposideridae). *Journal of Zoology*, **166**, 323–335.
- Ramasindrazana, B., Goodman, S. M., Rakotondramanana, C. F., & Schoeman, M. C. (2013). Morphological and echolocation call variation in Malagasy trident bats, *Triaenops Dobson*, 1871 (Chiroptera: Hipposideridae). *Acta Chiropterologica*, **15**, 431–439.
- Rautenbach, I. L., Schlitter, D., & Braack, L. (1984). New distributional records of bats for the Republic of South Africa, with special reference to the Kruger National Park. *Koedoe*, **27**, 131–135.
- Revilliod, P. (1914). Les mammifères de la Nouvelle-Calédonie et des îles Loyalty. *Nova Caledonia Zoologie*, **1**, 342–365.
- Rocha, P. A., Tavares, V. D., Pedroso, M. A., Beltrao-Mendes, R., Ruiz-Esparza, J., & Ferrari, S. F. (2018). First record of *Dermanura anderseni* (Chiroptera, Phyllostomidae) for the Atlantic Forest. *Mammalia*, **82**, 388–392.
- Rodríguez-Herrera, B., Sanchez, R., & Pineda, W. (2011). First record of *Natalus lanatus* (Chiroptera: Natalidae) in Costa Rica, and current distribution of *Natalus* in the country. *Ecotropica*, **17**, 113–117.
- Rodríguez-San Pedro, A., Allendes, J. L., Castillo, M. L., Peñaranda, D. A., & Peña-Gómez, F. T. (2014). Distribution extension and new record of *Myotis atacamensis* (Lataste, 1892) (Chiroptera: Vespertilionidae) in Chile. *Check List*, **10**, 1164–1166.
- Rodríguez-San Pedro, A., & Simonetti, J. A. (2013). Acoustic identification of four species of bats (Order Chiroptera) in central Chile. *Bioacoustics*, **22**, 165–172.
- Ruedi, M., Csorba, G., Lin, L. K., & Chou, C. H. (2015). Molecular phylogeny and morphological revision of *Myotis* bats (Chiroptera: Vespertilionidae) from Taiwan and adjacent China. *Zootaxa*, **3920**, 301–342.
- Rui, A., Fabian, M., & Menegheti, J. (1999). Geographical distribution and morphological analysis of *Artibeus lituratus* Olfers and *Artibeus fimbriatus* Gray (Chiroptera, Phyllostomidae) in Rio Grande do Sul, Brazil. *Revista Brasileira de Zoologia*, **16**, 447–460.
- Salsamendi, E., Aihartza, J., Goiti, U., Almenar, D., & Garin, I. (2006). Echolocation calls and morphology in the Mehelyi's (*Rhinolophus mehelyi*) and Mediterranean (*R. euryale*) horseshoe bats: Implications for resource partitioning. *Hystrix, the Italian Journal of Mammalogy*, **16**, 149–158.
- Santini, L., González-Suárez, M., Russo, D., Gonzalez-Voyer, A., von Hardenberg, A., & Ancillotto, L. (2019). One strategy does not fit all: Determinants of urban adaptation in mammals. *Ecology Letters*, **22**, 365–376.
- Scanlon, A., Petit, S., & Bottroff, G. (2014). The conservation status of bats in Fiji. *Oryx*, **48**, 451–459.
- Sedlock, J. L., Kruger, F., & Clare, E. L. (2014). Island bat diets: Does it matter more who you are or where you live? *Molecular Ecology*, **23**, 3684–3694.

- Sedlock, J. L., & Weyandt, S. E. (2009). Genetic divergence between morphologically and acoustically cryptic bats: Novel niche partitioning or recent contact? *Journal of Zoology*, **279**, 388–395.
- Shahbaz, M., Javid, A., Hussain, S. M., Ashraf, M., & Azmat, H. (2015). Recent Record of desert yellow house bat, *Scotoecus palidus* (Order Chiroptera) from Punjab, Pakistan. *Journal of Animal and Plant Sciences*, **25**, 599–602.
- Silvis, A., Perry, R., & Ford, W. M. (2016). Relationships of three species of bats impacted by white-nose syndrome to forest condition and management. *General Technical Report SRS-214*. Asheville, NC: U.S. Department of Agriculture Forest Service, Southern, **214**, 1–48.
- Tavares, V. C. (2013). Phyllostomid bat wings from Atlantic Forest bat ensembles: An ecomorphological study. *Chiroptera Neotropical*, **19**, 57–70.
- Taylor, P. J. (1999). Echolocation calls of twenty southern African bat species. *South African Journal of Zoology*, **34**, 114–124.
- Troughton, E. L. G. (1931). Three new bats of the genera *Pteropus*, *Nyctimene* and *Chaerephon* from Melanesia. *Proceedings of the Linnean Society of New South Wales*, **56**, 204–209.
- Trtkovic, N., Pavlinic, I., & Haring, E. (2005). Four species of long-eared bats (*Plecotus*, Geoffroy, 1818; Mammalia, Vespertilionidae) in Croatia: Field identification and distribution. *Folia Zoologica*, **54**, 75–88. 146.
- Vehrencamp, S. L., Stiles, F. G., & Bradbury, J. W. (1977). Observations on the foraging behavior and avian prey of the neotropical carnivorous bat, *Vampyrus spectrum*. *Journal of Mammalogy*, **58**(4), 469–478. <https://doi.org/10.2307/1379995>
- Velazco, P. M., & Lim, B. K. (2014). A new species of broad-nosed bat *Platyrrhinus* Saussure, 1860 (Chiroptera: Phyllostomidae) from the Guianan Shield. *Zootaxa*, **3796**, 175–193.
- Vieira, T. B., Mendes, P., Oprea, M., & Ditchfield, A. D. (2009). New record of *Lasiurus blossevillei* (Chiroptera: Vespertilionidae) for the restinga ecosystem, and second occurrence for the state of Espírito Santo, southeastern Brazil. *Mammalia*, **73**, 155–157.
- Voigt, C. C., & Lewanzik, D. (2012). 'No cost of echolocation for flying bats' revisited. *Journal of Comparative Physiology B*, **182**, 831–840.
- Webala, P. W., Rydell, J., Dick, C. W., Musila, S., & Patterson, B. D. (2019). Echolocation calls of high duty-cycle bats (Hipposideridae and Rhinonycteridae) from Kenya. *Journal of Bat Research & Conservation*, **12**, 10–20.
- Whitmore, N., Aplin, K., Arihafa, A., Armstrong, K., Cuthbert, R., Müller, C., Novera, J., Richards, S. J., Tamarua, W., Theischinger, G., & Venter, F. (2015). *A rapid biodiversity survey of Papua New Guinea's Manus and Mussau Islands*. Wildlife Conservation Society, Papua New Guinea Program.
- William, L., Richard, E., Tagide, N., & Michael, J. (2001). Pinnae and echolocation call differences between *Myotis californicus* and *M. ciliolabrum* (Chiroptera: Vespertilionidae). *Acta Chiropterologica*, **3**, 77–91.
- Williams, S. L., Willig, M. R., & Reid, F. A. (1995). Review of the *Tonatia bidens* complex (Mammalia: Chiroptera), with descriptions of two new subspecies. *Journal of Mammalogy*, **76**, 612–626.
- Wilman, H., Belmaker, J., Simpson, J., De La Rosa, C., Rivadeneira, M. M., & Jetz, W. (2014). EltonTraits 1.0: Species-level foraging attributes of the world's birds and mammals. *Ecology*, **95**, 2027–2027.
- Wordley, C. F. R., Foui, E. K., Mudappa, D., Sankaran, M., & Altringham, J. D. (2014). Acoustic identification of bats in the southern Western Ghats, India. *Acta Chiropterologica*, **16**, 213–222.
- Young, R. A., & Ford, G. I. (2000). Bat fauna of a semi-arid environment in central western Queensland, Australia. *Wildlife Research*, **27**, 203–215.
- Zamora-Gutierrez, V., Lopez-Gonzalez, C., MacSwiney, M. C., Fenton, B., Jones, G., Kalko, E. K. V., Puechmaille, S. J., Stathopoulos, V., & Jones, K. E. (2016). Acoustic identification of Mexican bats based on taxonomic and ecological constraints on call design. *Methods in Ecology and Evolution*, **7**, 1082–1091.
- Zhang, L. B., Jones, G., Zhang, J. S., Zhu, G. J., Parsons, S., Rossiter, S. J., & Zhang, S. Y. (2009). Recent surveys of bats (Mammalia: Chiroptera) from China. I. Rhinolophidae and Hipposideridae. *Acta Chiropterologica*, **11**, 71–88.
- Zhang, S., Zhao, H., Feng, J., Sheng, L., Li, Z., & Wang, L. (2000). Echolocation calls of *Myotis frater* (Chiroptera: Hipposideridae) during search flight. *Chinese Science Bulletin*, **45**, 1690–1692.